Yeast Chromatin Spread

Growth and Spheroplasting:

- 1. Grow 10 ml of yeast cells to log phase
- 2. Harvest and wash once with spheroplasting solution (1.2 M sorbitol, 0.1 M KPO₄)
- 3. Resuspend cells in 1 ml of spheroplasting solution
- 4. add 5 microliter of β-Mercaptoethanol and 10 μl of fresh zymolase (5 mg/ml of 100T zymolase in 1.0 M sorbitol)
- 5. Incubate at 30°C for 60 min.
- 6. Check the spheroplasting under phase contrast microscope.
- 7. Once 80-90% of the cells are spheroplasted, stop spheroplasting by adding ice cold stop solution (0.1M MES, pH 6.4, 1mM EDTA, 0.5 mM MgCl₂, 1 M sorbitol).
- 8. Harvest the spheroplasts at 4^oC (2000 rpm for 3 min)
- 9. Gently resuspend the spheroplasts in 1 ml of ice cold stop solution.

Making spread on the Slides:

- 1. Put 20 μl of spheroplasts on a clean slide
- 2. Add 40 µl of paraformaldehyde solution (4% solution with 3.4% sucrose)
- 3. Mix gently by tilting the slide
- 4. Add 80 µl of 1% Lipsol
- 5. Mix gently by tilting the slide
- 6. Add 80 µl of paraformaldehyde solution again
- 7. Mix again
- 8. Use a tooth pick or a pipet tip to gently mix and spread the contents around the slide.
- 9. Let slides dry overnight at RT.

Immunofluorescence:

- 1. Wash slides with 2 ml of 0.4% photoflow-200 (from Kodak)
- 2. Submerge slides in 1 X PBS for 10 min.
- 3. Remove excess liquid and immediately add 100 μ l of blocking solution (10 mg / ml BSA in 1XPBS)
- 4. Incubate for 15 min at RT
- 5. Shake of blocking solution and add 50 μl of primary antibody. Just drop a coverslip on, and incubate in a humid chamber for 60 min at RT.
- 6. Rinse of primary antibody by submerging the slide in 1XPBS for 5 min.
- 7. Add 50 μ l of secondary antibody and incubate in a humid chamber for 60 min as in step 5.
- 8. Rinse of secondary antibody by submerging the slide in 1XPBS for 5 min.
- 9. Repeat step 8 once.
- 10. Submerge slides in DAPI (1 μ g/ml in 1XPBS) for 5 min.
- 11. Rinse DAPI off by rinsing slides in PBS
- 12. Mount slides with mounting solution.